

Late revenge of analogue insulins among T1D-patients?

I am concerned: B.Teupe, Bad Mergentheim, Germany

► To the Editor – Background

This data analysis was provoked by the plight of a young patient: 1999 type-1-diabetes became manifest in a girl born 1992. Starting with ICT: Actrapid® / Protaphane®, 1 year later additional Humalog®, since 2 / 2001 Semilente®, CSII with Humalog® since 8 / 2004, changing 2008 shortly to Apidra® + Insuman® Infusat and since 4 / 2010 Novorapid® - in summary: she got porcine, human and all available analogue insulin sorts. Since about 2005 she progressively developed daily hyperglycaemia (up to ca. 400 – 600 mg/dl, independent from insulin dosages, up to nearly 600 I.U. of insulin did not change anything) with several ketoacidosis comas, late postprandial severe hypoglycaemia with often hypoglycaemic comas mainly in the early morning several times the week. HbA_{1c} worsened from minimal 6.4 % to at last 10.3 % over years (normal: 4.2 - 6.1%), not absolute uncommon in puberty.

She has been treated in some specialised diabetic institutions with several procedures (educational programs, multiple injections therapy, CSII, CGMS, experimentally intravenous therapy, underwent psychological and psychiatric explorations, family therapy) with no ameliorating effect.

17 years old, she was referred to me with the question: Do you have any idea how to help her? I could confirm the diagnosis of type-1-diabetes (C-peptide negative), but also found MODY-3-genes and extreme amounts of insulin-antibodies (23 210 nU/mL, 81%-binding) and at first extreme amounts of insulin-receptor-antibodies, but the latter seems to be a methodological problem (free circulating IA-IRA-complexes). As “all” immune suppressive therapies failed (glucocorticoids, azathioprin, mycophenolat³, Rituximab), first in case of urgent need (4 sessions in about 6 weeks) she is now precautionary constantly treated by immune adsorption (reliable vascular access via Cimino-shunt similar to dialysis with a IgA-column, ADASorb®) 2 days a fortnight⁴. By extracting the IA-concentrations into the normal range, her total daily dose sinks to about 50 I.U./ d /57 kg and her glucose metabolism stabilizes near normality, but her antibody production persists. Now we wait if the I(A)As will decline - which has not happened the last 2 years - and look for a definitely less invasive therapy, if possible, to avoid bone marrow transplantation.

► Abstract

Objective (aim): To collect IA-data status among T1D-patients in relation to their prior and present used insulin sorts (compared with healthy persons, T2D- and MODY-patients)

Methods: Out of 291 consecutive participants from more than 4000 of my CSII-patients we actually draw up 277 complete data sets of measured IA with two different methods (percentage of binding and absolute amounts), last HbA_{1c} and collected data belonging to their insulin therapy.

Results: At a significance level $p < 0,05$ we found

- that significant more patients using analogue insulin produce IAs compared with human insulin users,
- significant higher IA-levels in all T1D-patients compared with healthy persons,
- significant and / or borderline significant higher IA-levels in the group of analogue users compared with human insulin users,
- in all cases significant higher IA-levels in the group of the aspart insulin users compared with human-insulin users, users of the other analogue insulins and users of insulin derived from animals.

Conclusion: The careless prescribing of insulin analogues must be reconsidered under these results. It needs to be investigated urgently whether through the beginning of and the conversion to the use of analogue insulins a dangerous development is provoked towards a difficulty to control blood sugar metabolism as a result of IAs. All clinical diabetologists should review and follow up their diabetics on their IA-status routinely.

► Abbreviations

BMI (body mass index)

C-peptide (connecting peptide)

CSII (continuously subcutaneous insulin infusion = insulin pump treatment)

CGMS (continuous glucose monitoring system)

HbA_{1c} (percentage of N-terminal glycosylated β -chains of haemoglobin A₁)

IA (insulin antibody)

IAA (insulin-auto-antibody)

ICT (intensified insulin therapy = multiple injections)

IgA (immunoglobulin class A)

i.U. (international Units)

m (male)

MODY (maturity onset diabetes in young people, here only type 2 and 3)

nU / mL (nano units per millilitre)

SD (standard deviation)

T1D (type-1-diabetes)

T2D (type-2-diabetes)

► Introduction

Besides the rare insulinoma long be known, there is growing knowledge in the last years to the rarer hyperinsulinaemic hypoglycaemia without insulinoma and not induced by artificial insulin injection as insulin autoimmune syndrome (Hirata disease^{1,2}) produced by insulin(receptor)-antibodies in non-diabetic and type-2-diabetic persons^{5,6,7,8}. Despite some methodological problems⁹ there exists some knowledge of insulin antibodies as auto-antibodies of type-1-diabetics¹⁰ (accompanying the β -cell-destruction) and as acquired binding antibodies (even allergic) provoked by insulin therapy itself in all diabetics. Purification of insulin in the 1970th and the introduction of human insulin 1982 should and could reduce problems with bovine and porcine insulin antibodies¹¹. Recently there are reports to the higher levels of antibodies to analogue insulin but without clinical opinion^{12,13}.

Affected by the course of the disease of the above mentioned young woman, I found more patients in my collective with similar although weaker but therapy relevant problems with high insulin antibodies. Therefore I check all my patients routinely since October 2012 for IA for a better understanding of metabolism problems. These are my alerting preliminary results – the investigation will be extended and followed up.

► Subjects, Materials and Methods

Participants:

From a cohort of 291 consecutive participants (13.10.12 – 24.1.13) we got 277 complete records: 245 T1D-patients (83 m), 241 on pump treatment,

2 within this group with additional MODY-Genes (2 m),

5 T2D-patients (4 m),

9 MODY-patients (6 m),

18 non diabetic persons (6 m) (= no insulin treatment)

Table 1: Characterisation of the participants

		animal in comb.*	T1D no analogues**	analogue in comb.***	T2D	MODY animal* human** analogue***	Healthy	
	number total	245 (241 CSII)			5 (2 analog, 2 human, 1 no insulin)	9	18	
number total all	277	133	58	185	5	0	6	18
male	99	47	18	64	4	0	3	5
female	178	86	40	121	1	0	3	1
age (y)	45,4	50,2	49,3	42,7	70,2	57	54,0	47,2
± SD	±15,4	± 11,6	± 14,9	± 15,3	± 6,5	± 13,9	± 10,3	± 12,6
min - max	6-78	16-77	12-75	6-77	60-78	0	39-76	39-71
BMI	25,7	26,2	24,9	25,6	33,3	32,4	32,1	24,7
± SD	± 5,1	± 4,9	± 3,9	± 5,1	± 6,5	± 7,9	± 8,9	± 3,1
min - max	13,8-47,8	19,5-47,8	16,2-38,3	13,9-47,8	26,0-42,9	0	21,2-47,6	22,6-47,6
total daily dose (I.U./day)	46,8	46,6	44,1	46,3	41	94,7	68,2	
± SD	± 25,3	± 26,7	± 21,3	± 24,3	± 23,3	± 50,1	± 46,5	
min - max	12-160	12-160	12-100	15-160	15-60	0	34-150	25-150
diabetes duration (y)	26,5	35,3	33,1	24,4	26,6	30,2	24,0	
± SD	± 13,8	± 10,7	± 12,8	± 13,6	± 10,3	± 12,8	± 14,0	
min - max	1-67	11-67	8-67	1-56	12-37	0	17-48	4-44
insulin duration (y)	26,1	35,3	33,1	24,3	16	20,7	11,2	
± SD	± 13,9	± 10,8	± 12,9	± 13,6	± 8,5	± 13,8	± 6,4	
min - max	1-67	11-67	8-67	1-56	10-22	0	7-44	3-22
last HbA _{1c} in %	6,9	7,0	6,7	7,0	7,4	7,4	7,9	
± SD	± 1,0	± 1,0	± 0,7	± 1,0	± 1,0	± 1,2	± 1,3	
min - max	4,6-11,0	4,8-11,0	4,6-8,7	5,0-11,0	6,6-9,0	0	6,5-9,6	6,5-9,6
insulin binding in %	16,2	16,1	16,0	18,2	4,8	6,8	7,8	4,9
± SD	± 14,9	± 15,8	± 15,3	± 15,4	± 0,8	± 1,6	± 1,7	± 0,8
min - max	4-81	4-81	4-81	4-81	4-6	0	5-9	5-10
IA (nU/mL)	2151,8	1827,4	1789,3	2635,6	54,4	105,2	209,7	58,6
± SD	± 3682,4	± 3414,3	± 3234,6	± 4005,0	± 9,8	± 95,8	±246	± 16,7
min - max	50-23210	50-14902	50-14902	50-23210	50-72	0	50-296	50-637

* all patients treated prior with animal derived insulins (porcine and bovine), 2 present with porcine, with possible combination with other insulins, 37 without analogue contact (see Table 2)

** patients never treated with analogue insulins

***patients treated with analogues, possible contact with animal derived or human insulins

Table 2: Patients using animal insulin without analogues (2 present with porcine insulin) compared with patients using animal insulin and analogues too

	animal without analogues [± SD; min – max]	animal with analogues [± SD; min – max]
number total all	38	95
male	11	36
female	27	59
age (y)	54,6 [± 11,4; 16-75]	48,4 [± 11,3; 20-77]
BMI	24,9 [± 3,1; 20,0-31,3]	26,8 [± 5,3; 19,5-47,8]
total daily dose (I.U./day)	40,1 [± 17,6; 12-90]	49,3 [± 29,2; 15-160]
diabetes duration (y)	39,1 [± 10,3; 15-67]	33,8 [± 10,6; 11-56]
Insulin duration (y)	39,2 [± 10,4; 15-67]	33,7 [± 10,6; 11-56]
last HbA _{1c} in %	6,7 [± 0,7; 4,8-8,7]	7,0 [± 1,0; 5-11]
Insulin binding in %	15,0 [± 16,5; 4-81]	16,5 [± 15,6; 4-81]
IA (nU/mL)	1603,4 [± 3251,5; 50-14902]	1917 [± 3490,0; 50-23210]

Measurements and data collection:

- Insulin antibodies absolute (nU/mL) performed by MVZ Labor PD Dr. Volkmann und Kollegen GbR, Karlsruhe, Germany; [Normal range: < 110 nU/mL; gray zone: up to 250 nU/mL]

200 µL serum will be acidified with 250 µL 0,085M HCl, to dissociate immune complexes out of I(A)A und endogenous insulin. Afterwards add 100 µL of a mixture of activated carbon, dextran and methylcellulose in buffer A (Na-phosphate buffer with 0,33 M NaCl, 1,5% BSA), to bind endogenous insulin. After neutralization centrifuge the probes 10 min, 3750 x g. The supernatant solution will be used in the further radio immune precipitation assay. 100 µL of the supernatant solution will be diluted with 50 µL buffer A and 100 µL in buffer A diluted ¹²⁵I-Insulin (PerkinElmer, Rodgau). Parallel replace 50 µL buffer A by Insuman[®] rapid (Sanofi-Aventis, Frankfurt) for determination the unspecific binding of ¹²⁵I-Insulin. After incubation the probes overnight at 2-8°C, add 1,5 mL buffer B (0,1 M Tris-HCl, pH 8.0 with 17% PEG 6000). After further 20 min. incubation at 2-8°C centrifuge the probes (30 min., 3750 x g, 10°C). Add 1,5 mL buffer C (0,1 M Tris-HCl, pH 8.0 with 12,5% PEG 6000) to the aspirated supernatant solution and mix it thoroughly. After 20 min. of incubation at 2-8°C centrifuge the probes (30 min., 3750 x g, 10°C), aspirate the supernatant solution and measure the activity of the precipitated pellets in a gamma-counter. The difference (activities of the precipitates of the specific and the unspecific test) will be converted into nU/mL taking the date of calibration of ¹²⁵I-Insulin into account.

The test result as nU/mL is an absolute quantity, specificity and cross reactivity unclear.

- Percentage insulin binding levels by radio-immunoassay product of DIIAsource, formerly BioSource, Belgium (AIA RIA kit, KIP0091) performed by Laboratory Limbach Heidelberg Germany; [Normal range: < 10 %; borderline: 10 - 15 %]

The presence of circulating anti-insulin antibodies in patients treated with insulin will be assessed on a semi-quantitative basis by the determination of the binding of ¹²⁵I-Tyr-A₁₄-insulin in the serum fraction by the polyethylene glycol (PEG) (gammaglobulins) was precipitated. The used kit determined if the samples contain free anti-insulin antibodies, that are not bound to the insulin. The concentration of insulin in the tracer is 0.42 nmol / 100 µl of tracer. The origin of insulin is human recombinant with a purity > 99 % by HPLC ordered at Fitzgerald (USA). Item 1302129. The insulin is mono-iodated on position A14 and purified by HPLC. Intra- and inter-assay CVs: 1,6-20% (lower CVs in higher IA-concentrations); Mean of 80 healthy persons: 5,5 %; mean + 3 SD (= 8,2%): I(A)As positive.

This test does not measure the bound insulin in patients but in his serum in vitro under special conditions. The test result as percentage of binding is a relative number and not an absolute quantity, more than qualitative (antibody yes or no), less than quantitatively (= "semi quantitative"), specificity and cross reactivity unclear.

- The HbA_{1c}-measurements were mostly done with affinity chromatography (Afinion[™], product of Axis-Shield PoC AS, Oslo, Norway), mostly performed by ourselves. [Normal range: < 6 % (< 42,1 mmol/mol); coefficient of variation CV < 1,4%]
- Since October 2012 we extended our routine blood test for measuring the I(A)As in 2 different laboratories in all T1D-patients of our CSII-courses and ambulatory. We routinely asked them for their medical history. The data of age, gender, weight, height, diabetes duration, insulin treatment duration, total daily dose, all previous and present insulin sorts and their application duration, autoimmune state by the patients and their relatives, immune suppressive therapy (present and past) and last HbA_{1c} were filled in a questionnaire and in any doubt or obvious contradiction in the questionnaire we checked stored data and made interviews, using if necessary the services of medical reports. To control the details of the normal values of the laboratories, we invited all healthy escorts of the T1D-patients, T2D-patients and MODY-patients from our ambulatory to the same procedure.
- These data were transferred in an Excel data file (Microsoft[®], table calculation program), controlled by 2 persons and imported for statistical analyses into the statistic program Systat[®] 10.2 (www.systat.com), controlled by 3 persons, 2 of them are statistic professionals.
- As the data were not normal distributed we used the *Mann-Whitney U test* as a non-parametric statistical hypothesis test for assessing whether one of two samples of independent observations tends to have larger values than the other and the *Kruskal-Wallis one-way analysis of variance test* by ranks for testing whether samples originate from the

same distribution, if we compare more than two samples. With *Pearson X-squared test* we tested some null hypotheses. We define the limit for significance level to $p = 0,05$.

Descriptive statistics (mean values, standard deviations, correlations) were done by Excel, non parametric tests were performed by Systat®.

► Limitations, Possible Bias and Difficulties

For a better reminder, we gave our patients a list of about hundred historical and actual insulin product names, additionally it was easier for the patients to recollect some insulin producing companies, years of use and / or the origin of insulin. Therefore they could choose the origin of insulin (bovine, porcine, human, analogue) or special information as importations or foreign insulin. Many of the collected data were very precisely, p. e. reconstructed and confirmed by medical prescriptions. Sometimes it was easier for the patients to remind for some insulin producing companies, years of use and / or the origin of insulin – that enables reconstructions too. The duration of application was more imprecise, yet you could depend on semi quantitative results (shorter or longer than weeks, months, years). At all, we had to exclude only 4,8 % of patients due to lack of reliable data.

We could not discriminate between insulin auto-antibodies and acquired antibodies and non specificity due to cross reactivity between different insulin sorts. Though in average the influence of IAAs in our patients should be much smaller than in children cohorts, as their mean age was 45,4 years with an averaged diabetes duration of 26,5 years.

It is difficult to recruit “pure” groups of T1D-patients that use exclusively only one sort of insulin over a longer time – and for the future nearly impossible. Before introduction of human insulin about 30 years ago, they could have used only animal derived insulin (mostly bovine and porcine, purified in the 1970th). The first analogue insulin starts about 17 years ago. So the greatest majority has or has had contact with different sorts, different purities, different durations and past periods. Nowadays, there is a growing group treated only with analogues – but we have no peer group for comparison purposes. But we can form groups where one or some sorts are missing and compare them to each other. IA derived from animals may persist for more than 20 years¹¹.

Table 3: insulin usage groups in all of our patients

group	insulin sorts	number	
0	only regular	64	different products
1*	only aspart	80	Novorapid®
2	only glulisin	7	Apidra®
3	only lispro	53	Humalog®,Liprolog®
4*	aspart + lispro	35	
5	glulisin + lispro	4	
6*	aspart + lispro + glulisin	5	
sum		248	
significant different levels of IA absolute: $p = 0,003$, for %-binding $p = 0,004$, compared to group 0			

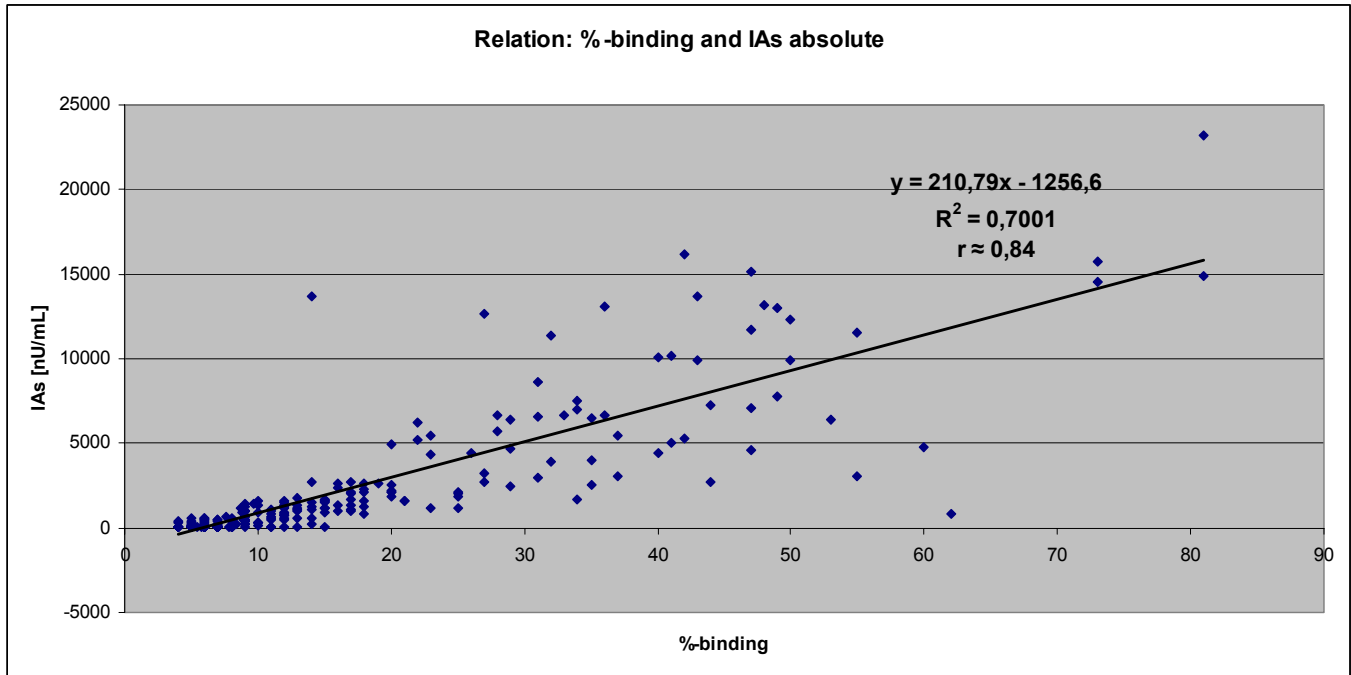
We selected our CSII-patients (241 of 245 T1D-patients) from our highly specialised institution with the peculiarity that the influence of any long acting insulin, the analogues, too, stopped mostly years ago totally.

As some of our significances failed sharply the 5 % level, we urgently need bigger groups. The power of these data suffers from the high dispersion (high standard deviation) of the values.

► Results

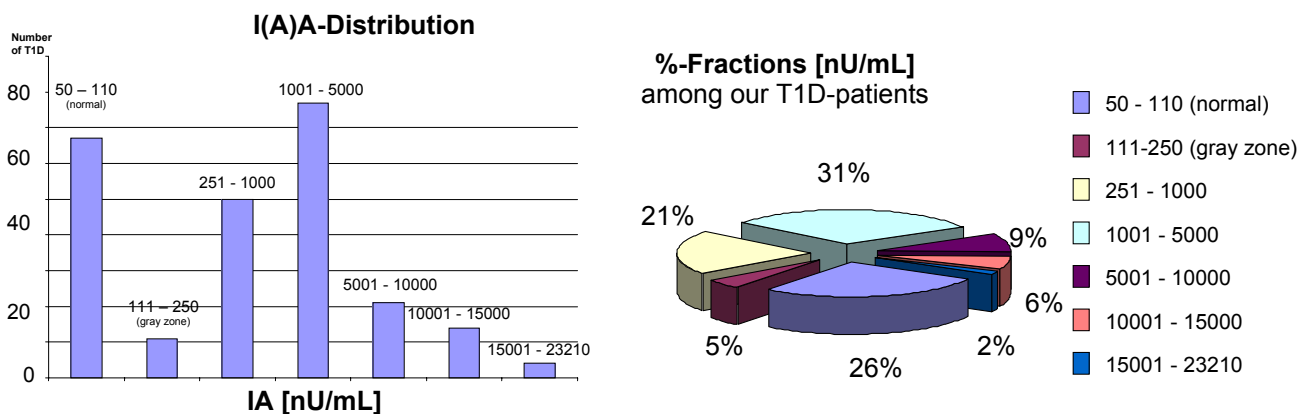
- Correlation between IA-binding and IA absolute is shown in the following figure. Repeated measuring of some of the runaway values showed similar differences. Therefore each of the compared methods have their specific problems, or they differ in what they are measuring. Probably these differences also reflect the semi quantitative and the absolute method.

Figure 1: Relationship of the percentage of bound insulin to the absolute amount of insulin antibodies [nU/mL]



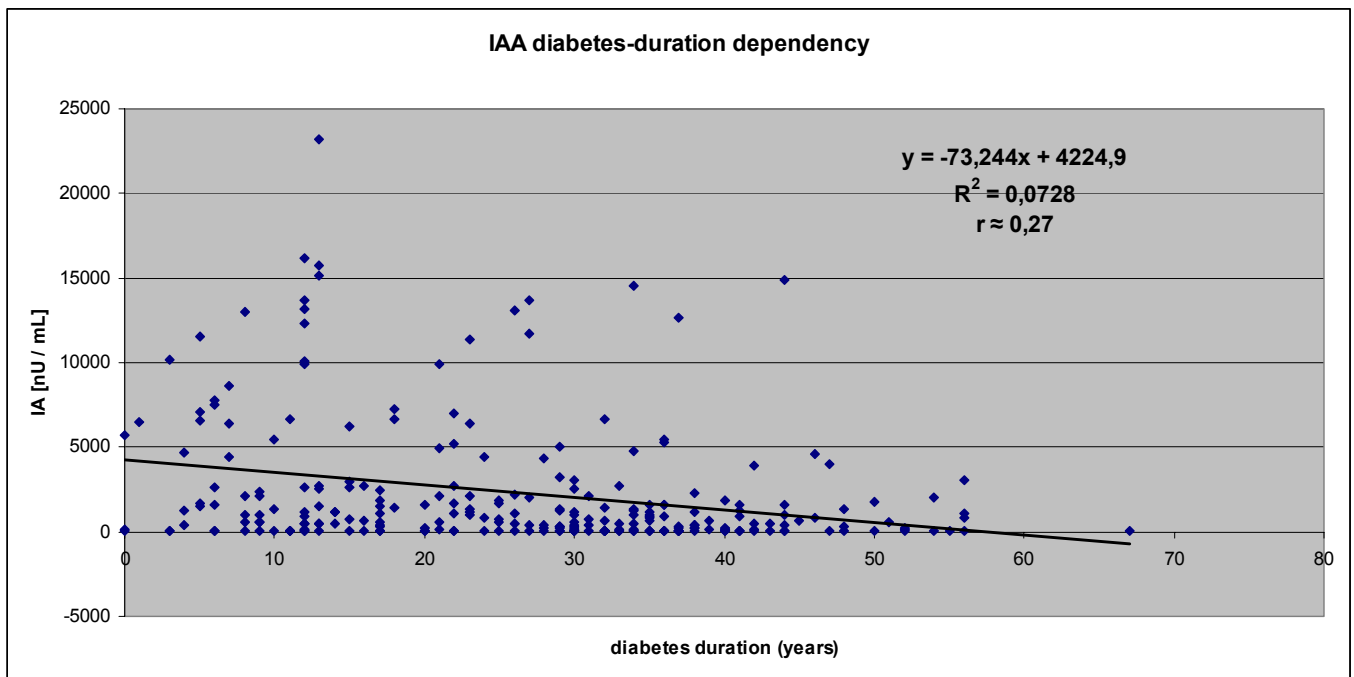
- Both IA-values for all healthy subjects were inside the normal ranges and confirmed these normal ranges for our whole collective.
- We found significant higher IA-levels in all T1D-patients compared with healthy persons ($p < 0,000$).
- 69 % of all T1D-patients exceed the IA normal range, 60 % the gray zone.

Figure 2: Distribution of IAs among T1D-patients



- Significant more patients (almost twice as many) using analogue insulin produce IAs ($p = 0,023$; $p = 0,04$) compared with human insulin users,
- significant higher IA-levels in the group of analogue users compared with the rest of the group ($p < 0,05$),
- borderline significant higher IA-levels in the group of analogue users compared with human insulin users ($p = 0,07$),
- in all cases significant higher IA-levels in the group of the aspart insulin users compared with human insulin users, users of the other analogue insulins and users of insulin derived from animals ($p = 0,003$).
- The group of T2D- and MODY-patients is too small to make strong statements. Maybe, the immunogenic impacts are less pronounced as rather an indication that autoimmune genes also favour the IAs.
- There was no correlation between HbA_{1c} and amount of IA due to the whole participants, but the HbA_{1c}-values of our problematic patients were higher.
- It exists a weak correlation between I(A)As and diabetes-duration with a declining tendency (see Figure 2). Other interesting correlations as to age, insulin and other autoimmune diseases were too weak or not examinable due to weak data.

Figure 3: Relationship between I(A)As and diabetes duration



► Discussion and Conclusions

What is fact, what can we hypothesize? Animal derived insulin and also analogue insulin are not produced naturally by the human body and all sorts are administered at unnatural routes.

Therefore the human immune system may detect and fend them off with humoral factors as it does it with other foreign proteins, too.

We have measured significant larger different titers of IAs between T1D-patients treated with short acting analogue insulin in contrast to T1D-patients treated with short acting human insulin (may be only the effect of insulin aspart). First, no more, no less. We can further conclude from our data that independent of high purity and origin, therapeutic insulins continue to be immunogenic in humans. Finding antibodies is not a normal state, and the extent is surprising - but it is initially only a laboratory and not (yet) a disease. We should ask: what do they matter? We measured IAs with 2 different methods: %-binding of insulin and absolute concentrations of IAs (nU/mL). In our data we see that they correlate indeed, but not very strongly ($r \approx 0,84$). In some persons there are big differences, therefore the methods have different problems or they do not testify the same: when measuring percentage of binding-insulin there is always given

the same aliquot to patients serum samples independent from the patients insulin demand (bigger differences in insulin sensitivity and need for eating and correction). But if we understand well the clinical symptoms of hyperglycaemia when eating with administered insulin and late postprandial hypoglycaemia as binding and releasing processes from the IA-buffer (due to the law of mass action), then this process in vivo is much more dynamic and not comparable with the % of bound insulin in vitro with constant insulin additive. If we look at clinical consequences (hyperglycaemia – hypoglycaemia) this data seems less relevant. If we want to describe the status of the IA-producing process, it is more reliable to determine absolute amounts or concentrations of IAs.

Even if the frequency and the amount of IAs may be biased in our cohort by several factors (p. e. nearly exclusive CSII-patients, no pure groups, high standard deviations,...), the phenomenon of potentially life-threatening high titers of IAs exists as shown definitely in one of them.

We see correlations and one is trying to take causalities. But we have to differentiate the first period of time of type-1-diabetes with insulin-auto-antibodies (IAA): they may climb to high values and may disappear relatively quickly and the possible parallel development later on or at the same time with acquired insulin-antibodies (IA, which cannot surely be differentiated specifically from the IAAs). But if there are significant differences between the frequency of the occurrence due to sort and / or the amounts of I(A)As between human insulin and analogue insulin, then at least these differences seem to be provoked by IAs.

Further on we see in our data that the more likely that T1D-patients were in contact with analogues the higher are the titers in nU/mL: 1603,4 for treated with animal derived insulin, without any contact to analogues - 1917 for animal derived insulin, with contact to analogues; 1789,3 no analogue contact - 2635,6 with analogue contact. In this sense we can break a lance(t) for porcine insulin.

When on the one hand pediatricians see T1D-patients, then they look at the beginning of T1D with different high titers of IAA. Maybe, in seldom cases they control them quantitatively when they disappear, but naturally they have no good chance to see the development of acquired IAs during the short period of the following part of the childhood. When on the other hand an internal specialist sees T1D-patients, he then gets the T1D-patients sent from pediatricians, is also visited by new diagnosed grown-up T1D-patients and sees the follow-up for decades in both groups – and therefore he is confronted with the increasing / decreasing IAAs and the increasing IAs. In a short period up to 2 years, Mianowska¹⁵ saw a climbing of the frequency of semi quantitatively I(A)As from 80 % to 97.9 % with no difference between the insulin sorts – that is no proof that those children with T1D will not be in danger to develop acquired IAs sort depending in bigger amounts later on. He sees this phenomenon as a consequence of the T1D autoimmune process or the immunogenic consequences of insulin therapy that would disappear spontaneously within 6 – 12 months. The diabetes duration of our problematic patients is much longer than 12 months. We detect more than 10000 nU/ml I(A)As in 16 of our 245 T1D-patients with diabetes duration longer than 10 years, in 8 longer than 20 years, in 3 longer than 30 years. Will they or some of them persist for a lifetime? This discussion remembered me on the disastrous discussion in the past that children will not develop late-complications.

We have no data of the influence of the long-term analogue insulins to IAs. The most of our patients never had and have contact with analogue long term insulin, or if so, only for a short time or a long time ago.

Unfortunately, we cannot predict the course of the further development of IA, we have no follow up yet. We assume that IAAs disappear more quickly and the acquired IAs will stay very long. If we weigh human and analogue short acting insulins due to the direct effect of bloodglucose and the practicability in the everyday life, we see advantages and disadvantages for both insulin sorts, likely in different patients and different life circumstances. Therefore, there is no need to principally start with analogues concerning the therapy. It is too simple to call it the price of modern therapy or modern lifestyle. But if you prescribe analogue insulin, you should inform the patient to this risk and control their IA-status routinely. As we proclaim “empowerment” and “informed decision”, we should demand warning notices in the enclosed package insert of insulin, if our findings should be confirmed.

Furthermore we should be more sensitive when we see daily long stretched hyperglycaemia followed by late postprandial severe hypoglycaemia, especially in the early morning, and take in

consideration IA if bigger total daily dosages of insulin occur and should investigate future surveillance of genesis and onset of acquired IA, their course and therapeutic impact. When Fineberg¹⁶ et al summarize: “Little proof exists, however, that the development of insulin antibodies (IAs) to exogenous insulin therapy affects integrated glucose control, insulin dose requirements, and incidence of hypoglycaemia...” then Ishizuka T et al.¹⁴ is cited “encountered two patients who developed daytime hyperglycaemia and early morning hypoglycaemia because of insulin antibody (IA) that the affinity was extremely lower and the capacity extremely higher than those of IA in the insulin autoimmune syndrome, after their insulin treatment were changed from human insulin to analogue insulin.”

And I encountered a young patient with type-1-diabetes, who can only survive by continuous immune adsorption therapy due to IA (maximum of 23210 nU/mL, 81-% binding) since more than 2 years, likely caused by analogue insulin, but not by aspart insulin (because she only has had contact with aspart insulin after the diagnosis of extreme I(A)A-values). At this extreme endpoint, we can proof the life-threatening clinical impact of I(A)As. In this case, no standard therapy exists. Nevertheless is there an analogy to the seldom autoimmune Hirata disease, first described by Hirata et al. 1970. He reported a patient with insulin autoimmune syndrome, followed 1994 by reports of 197 Japanese persons with the same disease^{1,2,5,6,8} and T2D-person⁷, now in T1D-patients?

But at least 3 further T1D-patients of my collective have bigger therapy-problems (daytime long stretched hyperglycaemia, nocturnal severe hypoglycaemia) probably due to high IA-levels (6373 nU/mL – 29 %; 16188 nU/mL – 42 %; 15178 nU/mL – 73 %) and some other patients with similar quantitative combinations of nU/mL and % of IAs have obviously no therapy problems. If the insulin-IA-associates have further negative aspects, we have even less informations. I fear that we look only at the tip of the iceberg – or did I and my colleagues systematically check our patients regarding to this phenomenon so far?

As an attempt to respond the title’s question, we have to extend it: Late revenge of analogue insulins and/or footprints of T1D autoimmune process and/or unavoidable immunogenic consequences of insulin therapy and/or Hirata-disease like among T1D-patients? Depending on the patient we look at the beginning of T1D more to the footprints of autoimmune process and later on more to unavoidable immunogenic consequences of insulin therapy, may be sometimes an analogy to the seldom Hirata-disease (associated with HLA-DRB1*0406/DQA1*0301/DQB1*0302?) – but we can not say exactly what of these options or mixtures exist in the individual case – any way is particularly troubling that one or more analogue insulin may play a triggering or reinforcing role. Now we can make mistakes in two directions: Optimistic appeasing or panic mongering. We have more anxious questions and should avoid a false sense of security. We should rather ask if there is an avalanche to come on the long term? Does this phenomenon remain an exception or how often will it occur? We should investigate it immediately – may be we have a chance to prevent it.

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► References

Literature research by PUBMED.DE, DIMDI, Literature-Appendix at publications, spontaneous

- 1 Hirata Y; Uchigata Y: Insulin autoimmune syndrome in Japan. *Diabetes research and clinical practice* 1994; 24 Suppl; p. S153-7
- 2 Uchigata Y , Eguchi Y, Takayama-Hasumi S, Omori : Insulin autoimmune syndrome (Hirata Disease): clinical features and epidemiology in Japan Review Article. *Diabetes Research and Clinical Practice* 1994, 22, Issues 2–3, p89-94
- 3 Segal, Allgrove et al.: Severe insulin resistance secondary to insulin antibodies: successful treatment with immunosuppressant MMF. *Pediatric Diabetes* 2008, 9, p. 250 - 254
- 4 Koyama R; Nakanishi K; Kato M; Yamashita S; Kuwahara H; Katori H: Hypoglycemia and hyperglycemia due to insulin antibodies against therapeutic human insulin: treatment with double filtration plasmapheresis and prednisolone. *The American journal of the medical sciences* 2005; 329 (5); p. 259-64
- 5 Yeung CW; Mak CM; Lam KS; Tam S: Hypoglycaemia due to autoimmune insulin syndrome in a 78-year-old Chinese man. *British journal of biomedical science* 2012; 69 (2); p. 80-82
- 6 Nasu T; Suzuki R; Okamoto Y; Miyata K; Uno A; Nakao R; Kawashima A; Nakao T; Kondo M: Late postprandial hypoglycemia due to bioactive insulin dissociation from autoantibody leading to unconsciousness in a patient with insulin autoimmune syndrome. *Internal medicine (Tokyo, Japan)* 2011; 50 (4); p. 339-43
- 7 Zhao TY; Li F; Xiong ZY: Frequent reoccurrence of hypoglycemia in a type 2 diabetic patient with insulin antibodies. *Molecular diagnosis & therapy* 2010; 14 (4); p. 237-41
- 8 Paiva ES; Pereira AE; Lombardi MT; Nishida SK; Tachibana TT; Ferrer C; Hauache OM; Vieira JG; Reis AF: Insulin autoimmune syndrome (Hirata disease) as differential diagnosis in patients with hyperinsulinemic hypoglycemia. *Pancreas* 2006; 32 (4); p. 431-2
- 9 Kim S; Yun YM; Hur M; Moon HW; Kim JQ: The effects of anti-insulin antibodies and crossreactivity with human recombinant insulin analogues in the E170 insulin immunometric assay. *The Korean journal of laboratory medicine* 2011; 31 (1); p. 22-9
- 10 Eisenbarth GS; Jeffrey J: The natural history of type 1A diabetes. *Arquivos brasileiros de endocrinologia e metabiologia* 2008; 52 (2); p. 146-55
- 11 Oak S; Phan TH; Gilliam LK; Hirsch IB; Hampe CS: Animal insulin therapy induces a biased insulin antibody response that persists for years after introduction of human insulin. *Acta diabetologica* 2010; 47 (2); p. 131-5
- 12 Klingensmith GJ: Insulin antibodies - are they still with us? Do they matter? *Pediatric diabetes* 2011; 12 (2); p. 75-7
- 13 Chen JW; Frystyk J; Lauritzen T; Christiansen JS: Impact of insulin antibodies on insulin aspart pharmacokinetics and pharmacodynamics after 12-week treatment with multiple daily injections of biphasic insulin aspart 30 in patients with type 1 diabetes. *European journal of endocrinology / European Federation of Endocrine Societies* 2005; 153 (6); p. 907-13
- 14 Ishizuka T; Ogawa S; Mori T; Nako K; Nakamichi T; Oka Y; Ito S: Characteristics of the antibodies of two patients who developed daytime hyperglycemia and morning hypoglycemia because of insulin antibodies. *Diabetes research and clinical practice* 2009; 84 (2); p. e21-3
- 15 Mianowska B; Szadkowska A; Pietrzak I; Zmyslowska A; Wegner O; Tomczonek J; Bodalski J; Mlynarski W: Immunogenicity of different brands of human insulin and rapid-acting insulin analogs in insulin-naive children with type 1 diabetes. *Pediatric diabetes* 2011; 12 (2); p. 78-84
- 16 Fineberg SE; Kawabata TT; Finco-Kent D; Fountaine RJ; Finch GL; Krasner AS: Immunological responses to exogenous insulin. *Endocrine reviews* (2007); 28 (6); p. 625-52